L Number	Hits	Search Text	DB	Time stamp
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2	0	((ttge or temporal adj1 temperature adj1 (gradient or gel) or temperature adj1 sweep)) same (t adj1 cell)	USPAT; US-PGPUB	2002/12/13 13:05
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=> s (ttge or temporal (w) temperature or temperature (w) sweep)
L1 200 (TTGE OR TEMPORAL (W) TEMPERATURE OR TEMPERATURE (W) SWEEP)

=> s l1 and T(w)cell
L2 7 L1 AND T(W) CELL

=> dup rem l2
PROCESSING COMPLETED FOR L2
L3 5 DUP REM L2 (2 DUPLICATES REMOVED)

=> d 1-5 bib ab

L3 ANSWER 1 OF 5 CAPLUS COPYRIGHT 2002 ACS
AN 2002:930975 CAPLUS
TI Microsatellite mutations of transforming growth factor-.beta. receptor type II and caspase-5 occur in human precursor **T-cell** lymphoblastic lymphomas/leukemias in vivo but are not associated with hMSH2 or hMLH1 promoter methylation
AU Scott, Stuart; Kimura, Tomofumi; Ichinohasama, Ryo; Bergen, Susan; Magliocco, Anthony; Reimer, Cara; Kerviche, Annette; Sheridan, David; DeCoteau, John F.
CS Royal University Hospital, Department of Pathology, Saskatoon Cancer Centre, University of Saskatchewan, Hospital Drive, Saskatoon, SK, 103, Can.
SO Leukemia Research (2003), 27(1), 23-34
CODEN: LEREDD; ISSN: 0145-2126
PB Elsevier Science Ltd.
DT Journal
LA English
AB In solid cancers, defective DNA mismatch repair (MMR) is most commonly caused by hMSH2 or hMLH1 mutations, or epigenetic silencing of hMLH1 by promoter hypermethylation, and results in the acquisition of characteristic frameshift microsatellite mutations of mononucleotide repeats located within the coding regions of defined target genes. We previously identified hMSH2 mutations in **T-cell** lymphoblastic lymphoma (T-LBL) patient tumor samples and others have reported coding region microsatellite mutations in **T-cell** acute lymphoblastic leukemia (T-ALL) cell lines. Thus, while MMR gene mutations are known to occur in some human T-lymphoblastic tumors in vivo, it is still unknown if the coding region microsatellite mutations detected in human cell lines also occur in vivo or if hMLH1 or hMSH2 promoter hypermethylation contributes to defective MMR in these tumors. We analyzed the TGF.beta.RII (A)10 and caspase-5 (A)10 coding region repeats in 16 human T-LBL/ALL patient tumor samples and identified six with microsatellite mutations in one or both repeats. There was no evidence of hMSH2 or hMLH1 promoter methylation as assessed by std. methylation specific PCR or by a novel temporal temp. gradient

electrophoresis (**TTGE**) method that analyzed 25 and 30 CpG sites in the hMLH1 and hMSH2 promoters, resp. Our results indicate that coding region microsatellite mutations characteristic of defective MMR occur in some human T-LBL/ALL in vivo but not as a consequence of hMLH1 or hMSH2 promoter hypermethylation. Furthermore, the identification of TGF.beta.RII and caspase-5 coding region mutations in vivo implicates these genes in the pathogenesis of human T-LBL/ALL.

L3 ANSWER 2 OF 5 MEDLINE DUPLICATE 1
AN 2001554807 MEDLINE
DN 21487560 PubMed ID: 11601137
TI Detection of clonal **T-cell** receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis.
AU Zhu D; Kadin M E; Samoszuk M
CS Nichols Institute, Quest Diagnostics, 33608 Ortega Highway, San Juan Capistrano, CA, USA.
SO AMERICAN JOURNAL OF CLINICAL PATHOLOGY, (2001 Oct) 116 (4) 527-34. Journal code: 0370470. ISSN: 0002-9173.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Abridged Index Medicus Journals; Priority Journals
EM 200110
ED Entered STN: 20011017
Last Updated on STN: 20011029
Entered Medline: 20011025
AB Limited combinatorial and junctional diversity in TCR-gamma gene rearrangement can result in amplification products that are difficult to interpret when analyzed by conventional gel electrophoresis methods that separate DNA based on size (polymerase chain reaction [PCR]/polyacrylamide gel electrophoresis [PAGE]). We describe a simple approach to the detection of clonal TCR-gamma gene rearrangement using **temporal temperature** gradient gel electrophoresis (**TTGE**) that uses a gradual and uniform increase in the temperature of a constant denaturing gel to resolve different DNA molecules based on base pair composition. We tested 42 clinical specimens (30 blood specimens and 12 formalin-fixed paraffin-embedded tissues) for **T-cell** clonality by PCR/PAGE and PCR/**TTGE**. Concordant results were obtained in only 22 specimens (52%). Of the 20 discordant cases, 18 samples were positive by **TTGE** and negative by PAGE. For all of the discordant cases, the **TTGE** yielded results that correlated better with the clinical data than did the PAGE method. We conclude that PCR/**TTGE** is more accurate and easier to perform than current methods for detecting clonal populations of T cells.

L3 ANSWER 3 OF 5 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:125303 BIOSIS
DN PREV200100125303
TI Detection of clonal **T-cell** receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis.
AU Zhu, D. (1); Samoszuk, M. (1)
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Laboratory Investigation, (January, 2001) Vol. 81, No. 1, pp. 184A. print. Meeting Info.: Annual Meeting of the United States and Canadian Academy of Pathology Atlanta, Georgia, USA March 03-09, 2001
ISSN: 0023-6837.
DT Conference
LA English
SL English

L3 ANSWER 4 OF 5 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:299406 BIOSIS
DN PREV200100299406
TI Detection of clonal **T-cell** receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis (**TTGE**.
AU Zhu, Dan (1); Samoszuk, Michael (1)
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Blood, (November 16, 2000) Vol. 96, No. 11 Part 1, pp. 127a. print.
Meeting Info.: 42nd Annual Meeting of the American Society of Hematology San Francisco, California, USA December 01-05, 2000 American Society of Hematology
. ISSN: 0006-4971.
DT Conference
LA English
SL English
AB The demonstration of a clonal **T-cell** receptor-gamma (TCR-gamma) gene rearrangement using polymerase chain reaction (PCR) followed by gel electrophoresis is a helpful tool for detecting neoplastic T-cells in tissues and blood. A significant limitation of this procedure, however, is the limited combinatorial and junctional diversity in TCR-gamma gene rearrangement which can result in amplification products that are difficult to interpret when analyzed by standard gel electrophoresis that separates DNA molecules based solely on size. We describe a simple new approach to the detection of clonal TCR-gamma gene rearrangement using **temporal temperature** gradient gel electrophoresis (**TTGE**) that can resolve DNA molecules with a difference of as little as a single base pair substitution. The new method employs a gradual and uniform increase in temperature of a constant denaturing gel that is much easier to prepare and use than present equivalent methods in clinical diagnostic laboratories. In this study, we analyzed 42 clinical samples of known or suspected **T-cell** malignancy (30 peripheral blood specimens and 12 formalin-fixed tissues) by the standard method and by PCR/**TTGE**. Concordant results were obtained in 34 specimens (81%). There were 6 cases that were positive by **TTGE** and negative by the standard method, and 2 that were positive by the standard method and negative by **TTGE**. For all specimens, the **TTGE** results were much easier to interpret than the standard method. Our data, therefore, suggest that **TTGE** is a more sensitive and specific method for detecting clonal populations of T-cells in fresh and formalin-fixed tissues than methods that rely on separation of amplicons based on size alone.

L3 ANSWER 5 OF 5 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 1999:161061 BIOSIS
DN PREV199900161061
TI Detection of **T-cell** receptor-gamma gene rearrangement by **temporal temperature** gradient electrophoresis (**TTGE**.
AU Cosar, E.; Alkan, S.
CS Dep. Pathol., Loyola Univ. Med. Cent., Maywood, IL USA
SO Modern Pathology, (Jan., 1999) Vol. 12, No. 1, pp. 134A.
Meeting Info.: Annual Meeting of the United States and Canadian Academy of Pathology San Francisco, California, USA March 20-26, 1999
ISSN: 0893-3952.
DT Conference
LA English

=> d his

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L1 200 S (TTGE OR TEMPORAL (W) TEMPERATURE OR TEMPERATURE (W) SWEEP)
L2 7 S L1 AND T(W)CELL
L3 5 DUP REM L2 (2 DUPLICATES REMOVED)

=> d l1 and clonal?
'AND' IS NOT A VALID FORMAT
'CLONAL?' IS NOT A VALID FORMAT
In a multifile environment, a format can only be used if it is valid
in at least one of the files. Refer to file specific help messages
or the STNGUIDE file for information on formats available in
individual files.
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=> s l1 and clonal?
L4 5 L1 AND CLONAL?

=> dup rem l4
PROCESSING COMPLETED FOR L4
L5 3 DUP REM L4 (2 DUPLICATES REMOVED)

=> d 1-3 bib ab

L5 ANSWER 1 OF 3 MEDLINE DUPLICATE 1
AN 2001554807 MEDLINE
DN 21487560 PubMed ID: 11601137
TI Detection of **clonal** T-cell receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis.
AU Zhu D; Kadin M E; Samoszuk M
CS Nichols Institute, Quest Diagnostics, 33608 Ortega Highway, San Juan Capistrano, CA, USA.
SO AMERICAN JOURNAL OF CLINICAL PATHOLOGY, (2001 Oct) 116 (4) 527-34.
Journal code: 0370470. ISSN: 0002-9173.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Abridged Index Medicus Journals; Priority Journals
EM 200110
ED Entered STN: 20011017
Last Updated on STN: 20011029
Entered Medline: 20011025
AB Limited combinatorial and junctional diversity in TCR-gamma gene rearrangement can result in amplification products that are difficult to interpret when analyzed by conventional gel electrophoresis methods that separate DNA based on size (polymerase chain reaction [PCR]/polyacrylamide gel electrophoresis [PAGE]). We describe a simple approach to the detection of **clonal** TCR-gamma gene rearrangement using **temporal temperature** gradient gel electrophoresis (**TTGE**) that uses a gradual and uniform increase in the temperature of a constant denaturing gel to resolve different DNA molecules based on base pair composition. We tested 42 clinical specimens (30 blood specimens and 12 formalin-fixed paraffin-embedded tissues) for T-cell **clonality** by PCR/PAGE and PCR/**TTGE**. Concordant results were obtained in only 22 specimens (52%). Of the 20 discordant cases, 18 samples were positive by **TTGE** and negative by PAGE. For all of the discordant cases, the **TTGE** yielded results that correlated better with the clinical data than did the PAGE method. We conclude that PCR/**TTGE** is more accurate and easier to perform than current methods for detecting **clonal** populations of T cells.

L5 ANSWER 2 OF 3 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:125303 BIOSIS
DN PREV200100125303
TI Detection of **clonal** T-cell receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis.
AU Zhu, D. (1); Samoszuk, M. (1)
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Laboratory Investigation, (January, 2001) Vol. 81, No. 1, pp. 184A. print.
Meeting Info.: Annual Meeting of the United States and Canadian Academy of Pathology Atlanta, Georgia, USA March 03-09, 2001
ISSN: 0023-6837.
DT Conference
LA English
SL English

L5 ANSWER 3 OF 3 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:299406 BIOSIS
DN PREV200100299406
TI Detection of **clonal** T-cell receptor-gamma gene rearrangement by PCR/**temporal temperature** gradient gel electrophoresis (**TTGE**.
AU Zhu, Dan (1); Samoszuk, Michael (1)
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Blood, (November 16, 2000) Vol. 96, No. 11 Part 1, pp. 127a. print.
Meeting Info.: 42nd Annual Meeting of the American Society of Hematology San Francisco, California, USA December 01-05, 2000 American Society of Hematology
. ISSN: 0006-4971.
DT Conference
LA English
SL English
AB The demonstration of a **clonal** T-cell receptor-gamma (TCR-gamma) gene rearrangement using polymerase chain reaction (PCR) followed by gel electrophoresis is a helpful tool for detecting neoplastic T-cells in tissues and blood. A significant limitation of this procedure, however, is the limited combinatorial and junctional diversity in TCR-gamma gene rearrangement which can result in amplification products that are difficult to interpret when analyzed by standard gel electrophoresis that separates DNA molecules based solely on size. We describe a simple new approach to the detection of **clonal** TCR-gamma gene rearrangement using **temporal temperature** gradient gel electrophoresis (**TTGE**) that can resolve DNA molecules with a difference of as little as a single base pair substitution. The new method employs a gradual and uniform increase in temperature of a constant denaturing gel that is much easier to prepare and use than present equivalent methods in clinical diagnostic laboratories. In this study, we analyzed 42 clinical samples of known or suspected T-cell malignancy (30 peripheral blood specimens and 12 formalin-fixed tissues) by the standard method and by PCR/**TTGE**. Concordant results were obtained in 34 specimens (81%). There were 6 cases that were positive by **TTGE** and negative by the standard method, and 2 that were positive by the standard method and negative by **TTGE**. For all specimens, the **TTGE** results were much easier to interpret than the standard method. Our data, therefore, suggest that **TTGE** is a more sensitive and specific method for detecting **clonal** populations of T-cells in fresh and formalin-fixed tissues than methods that rely on separation of amplicons based on size alone.

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FULL ESTIMATED COST                                        27.75      28.38

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FULL ESTIMATED COST                                         0.06      28.44

DISCOUNT AMOUNTS (FOR QUALIFYING ACCOUNTS)              SINCE FILE      TOTAL
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CA SUBSCRIBER PRICE                                         0.00      -0.62

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=> s samoszuk?/au and zhu?/au
L6            5 SAMOSZUK?/AU AND ZHU?/AU

=> dup rem l6
PROCESSING COMPLETED FOR L6
L7            3 DUP REM L6 (2 DUPLICATES REMOVED)

=> d 1-3 bib ab

L7   ANSWER 1 OF 3     MEDLINE                          DUPLICATE 1
AN   2001554807     MEDLINE
DN   21487560   PubMed ID: 11601137
TI   Detection of clonal T-cell receptor-gamma gene rearrangement by
     PCR/temporal temperature gradient gel electrophoresis.
AU   Zhu D; Kadin M E; Samoszuk M
CS   Nichols Institute, Quest Diagnostics, 33608 Ortega Highway, San Juan
     Capistrano, CA, USA.
SO   AMERICAN JOURNAL OF CLINICAL PATHOLOGY, (2001 Oct) 116 (4) 527-34.
     Journal code: 0370470. ISSN: 0002-9173.
CY   United States
DT   Journal; Article; (JOURNAL ARTICLE)
LA   English
FS   Abridged Index Medicus Journals; Priority Journals
EM   200110
ED   Entered STN: 20011017
     Last Updated on STN: 20011029
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Entered Medline: 20011025

AB Limited combinatorial and junctional diversity in TCR-gamma gene rearrangement can result in amplification products that are difficult to interpret when analyzed by conventional gel electrophoresis methods that separate DNA based on size (polymerase chain reaction [PCR]/polyacrylamide gel electrophoresis [PAGE]). We describe a simple approach to the detection of clonal TCR-gamma gene rearrangement using temporal temperature gradient gel electrophoresis (TTGE) that uses a gradual and uniform increase in the temperature of a constant denaturing gel to resolve different DNA molecules based on base pair composition. We tested 42 clinical specimens (30 blood specimens and 12 formalin-fixed paraffin-embedded tissues) for T-cell clonality by PCR/PAGE and PCR/TTGE. Concordant results were obtained in only 22 specimens (52%). Of the 20 discordant cases, 18 samples were positive by TTGE and negative by PAGE. For all of the discordant cases, the TTGE yielded results that correlated better with the clinical data than did the PAGE method. We conclude that PCR/TTGE is more accurate and easier to perform than current methods for detecting clonal populations of T cells.

L7 ANSWER 2 OF 3 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:125303 BIOSIS
DN PREV200100125303
TI Detection of clonal T-cell receptor-gamma gene rearrangement by PCR/temporal temperature gradient gel electrophoresis.
AU **Zhu, D. (1); Samoszuk, M. (1)**
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Laboratory Investigation, (January, 2001) Vol. 81, No. 1, pp. 184A. print.
Meeting Info.: Annual Meeting of the United States and Canadian Academy of Pathology Atlanta, Georgia, USA March 03-09, 2001
ISSN: 0023-6837.
DT Conference
LA English
SL English

L7 ANSWER 3 OF 3 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 2001:299406 BIOSIS
DN PREV200100299406
TI Detection of clonal T-cell receptor-gamma gene rearrangement by PCR/temporal temperature gradient gel electrophoresis (TTGE.
AU **Zhu, Dan (1); Samoszuk, Michael (1)**
CS (1) Nichols Institute, Quest Diagnostics, Inc., San Juan Capistrano, CA USA
SO Blood, (November 16, 2000) Vol. 96, No. 11 Part 1, pp. 127a. print.
Meeting Info.: 42nd Annual Meeting of the American Society of Hematology San Francisco, California, USA December 01-05, 2000 American Society of Hematology
. ISSN: 0006-4971.
DT Conference
LA English
SL English
AB The demonstration of a clonal T-cell receptor-gamma (TCR-gamma) gene rearrangement using polymerase chain reaction (PCR) followed by gel electrophoresis is a helpful tool for detecting neoplastic T-cells in tissues and blood. A significant limitation of this procedure, however, is the limited combinatorial and junctional diversity in TCR-gamma gene rearrangement which can result in amplification products that are difficult to interpret when analyzed by standard gel electrophoresis that separates DNA molecules based solely on size. We describe a simple new approach to the detection of clonal TCR-gamma gene rearrangement using temporal temperature gradient gel electrophoresis (TTGE) that can resolve DNA molecules with a difference of as little as a single base pair

substitution. The new method employs a gradual and uniform increase in temperature of a constant denaturing gel that is much easier to prepare and use than present equivalent methods in clinical diagnostic laboratories. In this study, we analyzed 42 clinical samples of known or suspected T-cell malignancy (30 peripheral blood specimens and 12 formalin-fixed tissues) by the standard method and by PCR/TTGE. Concordant results were obtained in 34 specimens (81%). There were 6 cases that were positive by TTGE and negative by the standard method, and 2 that were positive by the standard method and negative by TTGE. For all specimens, the TTGE results were much easier to interpret than the standard method. Our data, therefore, suggest that TTGE is a more sensitive and specific method for detecting clonal populations of T-cells in fresh and formalin-fixed tissues than methods that rely on separation of amplicons based on size alone.

=> s t(w)cell and multiple (4a) lesion#
L1 323 T(W) CELL AND MULTIPLE (4A) LESION#

=> s l1 and clonal?
L2 31 L1 AND CLONAL?

=> dup rem l2
PROCESSING COMPLETED FOR L2
L3 16 DUP REM L2 (15 DUPLICATES REMOVED)

=> d 1-16 ti

L3 ANSWER 1 OF 16 MEDLINE DUPLICATE 1
TI Plasmacytoma with aberrant expression of myeloid markers, **T-cell** markers, and cytokeratin.

L3 ANSWER 2 OF 16 MEDLINE
TI Anetoderma arising in cutaneous B-cell lymphoproliferative disease.

L3 ANSWER 3 OF 16 MEDLINE DUPLICATE 2
TI Accumulation of common **clonal** T cells in **multiple lesions** of sarcoidosis.

L3 ANSWER 4 OF 16 MEDLINE DUPLICATE 3
TI **Clonal** expansions of CD8(+) T cells dominate the **T cell** infiltrate in active **multiple** sclerosis **lesions** as shown by micromanipulation and single cell polymerase chain reaction.

L3 ANSWER 5 OF 16 MEDLINE DUPLICATE 4
TI In situ **T cell** responses against melanoma comprise high numbers of locally expanded **T cell** clonotypes.

L3 ANSWER 6 OF 16 MEDLINE DUPLICATE 5
TI Natural killer cell-derived large granular lymphocyte lymphoma of lung developed in a patient with hypersensitivity to mosquito bites and reactivated Epstein-Barr virus infection.

L3 ANSWER 7 OF 16 MEDLINE DUPLICATE 6
TI The dominant **T cell** clone is present in **multiple** regressing skin **lesions** and associated **T cell** lymphomas of patients with lymphomatoid papulosis.

L3 ANSWER 8 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
TI Detection of the same dominant **T cell** clone in **multiple** lymphomatoid papulosis (LyP) **lesions** and associated lymphomas.

L3 ANSWER 9 OF 16 MEDLINE DUPLICATE 7
TI Gamma delta **T cell** receptor analysis supports a role for HSP 70 selection of lymphocytes in **multiple** sclerosis **lesions**.

L3 ANSWER 10 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.DUPLICATE 8
TI Multiple sclerosis: Limited diversity of the V-delta-2-J-delta-3 **T-cell** receptor in chronic active lesions.

L3 ANSWER 11 OF 16 MEDLINE
TI Cutaneous follicular lymphoid hyperplasia with monotypic plasma cells. A

clinicopathologic study of 18 patients.

L3 ANSWER 12 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
TI Analyses of **T cell clonality** in **multiple** sialoadenitis **lesions** of IQI/Jcl mice.

L3 ANSWER 13 OF 16 MEDLINE DUPLICATE 9
TI Gamma delta **T cell** receptor repertoire in brain **lesions** of patients with **multiple** sclerosis.

L3 ANSWER 14 OF 16 MEDLINE DUPLICATE 10
TI Gamma delta **T-cell** receptor repertoire in acute **multiple** sclerosis **lesions**.

L3 ANSWER 15 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
TI **T CELL** SUBSETS AND LIPID MACROPHAGES IN **MULTIPLE** SCLEROSIS **LESIONS** IN-SITU CHARACTERIZATION USING MONO **CLONAL** ANTIBODIES.

L3 ANSWER 16 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
TI IDENTIFICATION AND DYNAMICS OF **T CELL** SUBSETS AND B CELLS DURING THE DEVELOPMENT OF **MULTIPLE** SCLEROSIS **LESIONS**.

=> d 7, 8 bib ab

L3 ANSWER 7 OF 16 MEDLINE DUPLICATE 6
AN 96183551 MEDLINE
DN 96183551 PubMed ID: 8618007
TI The dominant **T cell** clone is present in **multiple** regressing skin **lesions** and associated **T cell** lymphomas of patients with lymphomatoid papulosis.
AU Chott A; Vonderheid E C; Olbricht S; Miao N N; Balk S P; Kadin M E
CS Department of Pathology, Beth Israel Hospital, Boston, Massachusetts, USA.
NC RO1-CA 54062 (NCI)
SO JOURNAL OF INVESTIGATIVE DERMATOLOGY, (1996 Apr) 106 (4) 696-700.
Journal code: 0426720. ISSN: 0022-202X.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199606
ED Entered STN: 19960620
Last Updated on STN: 19960620
Entered Medline: 19960613
AB This study was undertaken to determine the **clonality** of lymphomatoid papulosis (LyP), its **clonal** relationship to lymphomas, which occur at high frequency in LyP patients, and to define the cell lineage of Reed-Sternberg-like cells in type A lesions of LyP. Punch biopsies of skin of 11 adult patients with LyP were analyzed for morphologic subtype of LyP, surface antigens, and **clonal** **T-cell** receptor (TCR) gene rearrangements. **Clonal** rearrangements were identified by semiquantitative polymerase chain reaction amplification and sequencing of TCR-beta chain genes in nine patients and TCR-gamma chain genes in two patients. A single dominant clone was detected in **multiple** separate LyP **lesions**, often of different histologies, in nine patients. The same clone was detected in LyP lesions and the anaplastic large cell lymphoma (ALCL) of 2 patients and the mycosis fungoides (MF) of 2 other patients. No dominant clone could be detected in one patient with LyP

uncomplicated by lymphoma or in a second patient with LyP and MF. A **T-cell** lineage was evident for RS-like cells in cell culture and in type A **lesions**. These results show that **multiple** regressing skin **lesions** and associated **T cell** lymphomas (MF and ALCL) are **clonally** related in most LyP patients, which suggest that the disease in these patients was initiated by a non-random genetic event.

L3 ANSWER 8 OF 16 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 1996:203303 BIOSIS
DN PREV199698759432
TI Detection of the same dominant **T cell** clone in **multiple** lymphomatoid papulosis (LyP) **lesions** and associated lymphomas.
AU Chott, A. (1); Vonderheid, E. C.; Miao, N.-N.; Balk, S. P.; Kadin, M. E.
CS (1) Beth Israel Hosp., Boston, MA USA
SO Modern Pathology, (1996) Vol. 9, No. 1, pp. 109A.
Meeting Info.: 1996 Annual Meeting of the United States and Canadian Academy of Pathology Washington, D.C., USA March 23-29, 1996
ISSN: 0893-3952.
DT Conference
LA English